

Molecular Mechanisms for Epigenetic Transcriptional Poised Memory State

Bethany Sump, Agustina D'Urso, Jason Brickner
Department of Molecular Biosciences, Northwestern University, Evanston IL



The major mechanism by which cells adapt to changes in their environment is by altering transcription. In some cases, the response to a stimulus is affected by the prior experiences of the cell. One such phenomenon is called *epigenetic transcriptional memory*; cell populations that have previously experienced a particular stimulus show a faster or stronger response to this stimulus in the future. Through work in budding yeast and HeLa cells, this phenomenon has been shown to be characterized by a unique transcriptional memory state that persists for 4-8 generations.

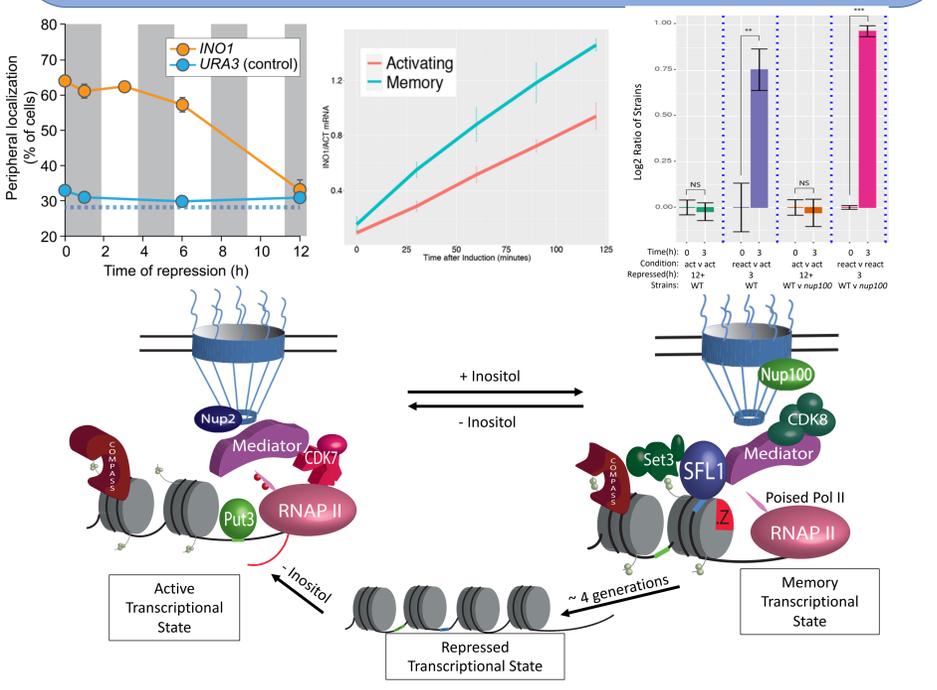


Figure 1: **INO1 Epigenetic Transcriptional Memory** A) Percent of cells that show *INO1* at the nuclear periphery post repression of activated cells. Gray to white bars indicate generations. B) Gene expression levels of *INO1* upon activation and reactivation with Opi1-AA (after 3 hours repression) by RTq-PCR. C) Competitive fitness benefit of populations undergoing reactivation by sequencing-based competition assay. D) Model for the different transcriptional states of the *INO1* gene.

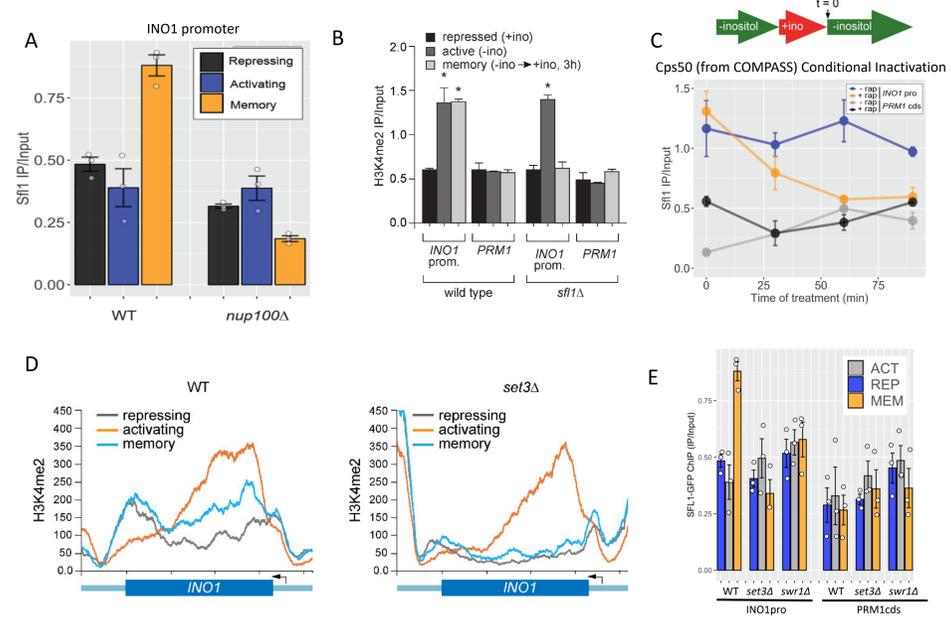


Figure 3: **The essential components of Epigenetic Transcriptional Memory form a feedback loop** A) Sfl1 ChIP at the *INO1* promoter region in WT and *nup100Δ* strains under repressing (+inositol), activating (-inositol), and memory (-inositol overnight, then 3 hours repressing) conditions. B) H3K4me2 ChIP in repressing, activating, and memory conditions (see above) at the *INO1* promoter in WT, *Sfl1Δ*, and *mrm1Δ* strains. C) ChIP-qPCR of Sfl1 at the *INO1* promoter region upon conditional inactivation of Cps50 by Anchor Away in memory conditions D) H3K4me2 ChIP-seq experiment focused over the *INO1* gene in WT and *set3Δ* strains. E) Sfl1-GFP ChIP-qPCR at the *INO1* promoter region in WT, *set3Δ*, and *swr1Δ* strains, showing that Sfl1 binding in memory conditions is dependent on chromatin modifiers.

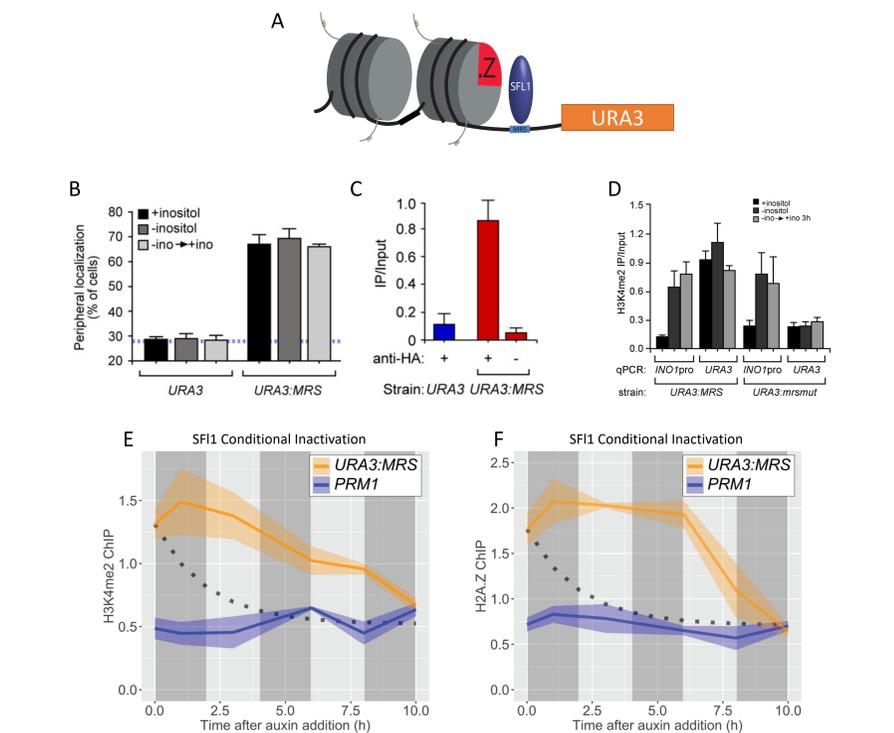


Figure 5: **Memory-specific chromatin marks are epigenetic when induced at ectopic locus by Memory Recruitment Sequence (MRS)** A) Model for integration of MRS at uninduced *URA3* locus. B) Percent of cells that show *URA3* at the nuclear periphery when in repressing (+inositol), activating (-inositol), and memory (overnight -inositol, then 3 hours repressing) conditions with or without MRS integration at *URA3*. C) ChIP-qPCR against HA-H2A.Z in strains either with or without MRS integrated at *URA3*. D) ChIP-qPCR against H3K4me2 at *INO1* promoter region and *URA3* under repressing, activating, and memory conditions (see above) when either the MRS or *mrm1Δ* is integrated at *URA3*. ChIP-qPCR against E) H3K4me2 or F) H2A.Z at *URA3* when the MRS is integrated at *URA3* after conditional inactivation of Sfl1 by Auxin-Induced Degrade (AID) technique.

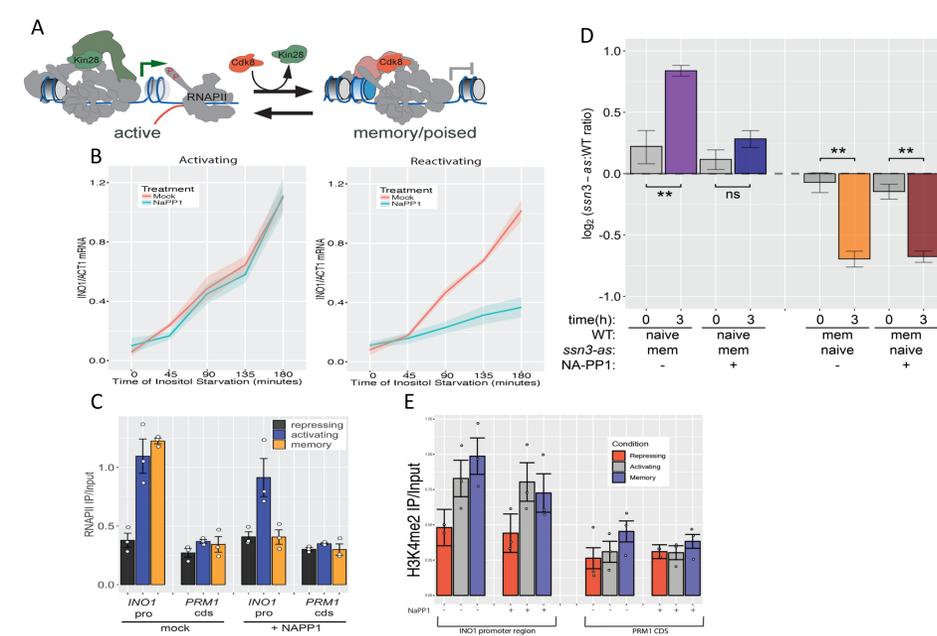


Figure 2: **Kinase Activity of Cdk8 is essential for RNA Polymerase II poising, but not chromatin changes** A) Model showing the remodeling of Mediator into the memory-specific state. B) RTq-PCR showing *INO1* mRNA levels during activating (left) and reactivating (right) transcriptional states when Cdk8 is catalytically active (red) or inactive (blue). C) ChIP-qPCR against RNA Polymerase II at the *INO1* promoter region under repressing (+inositol), activating (-inositol), and memory (overnight activating, 3 hours repressing) conditions, both while Cdk8 is active (mock) and inactive (+NAPP1). D) Sequencing-based competition assay showing the loss of the fitness benefit obtained by the 'reactivation' population if Cdk8 is inactive. E) ChIP-qPCR against H3K4me2 at the *INO1* promoter region under repressing, activating, and memory conditions (see above).

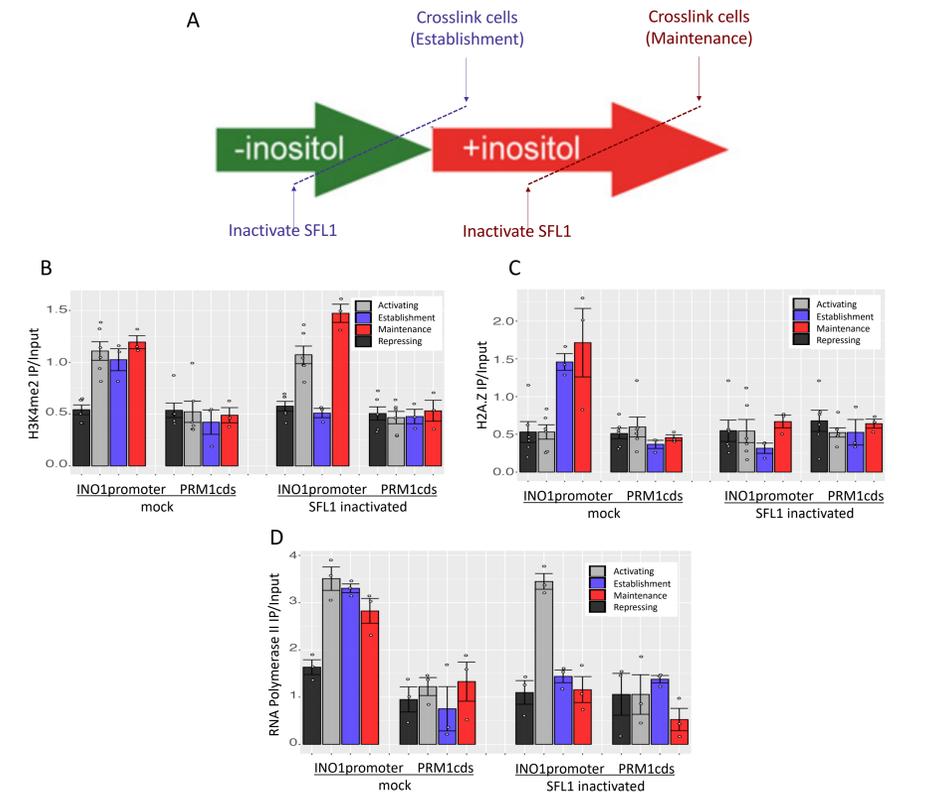


Figure 4: **Sfl1 plays roles in both the establishment and the maintenance of the memory-specific chromatin state** A) Schematic for investigating the separate roles of Sfl1 in the establishment and the maintenance of the memory-specific chromatin marks and presence of poised RNA Polymerase II. B) H3K4me2, C) H2A.Z, and D) RNA Polymerase II ChIP at the *INO1* promoter region in the conditions of repressing (+inositol), activating (-inositol), and 2 hours with Sfl1 removed either before (establishment) or after (maintenance) the switch into the memory state.

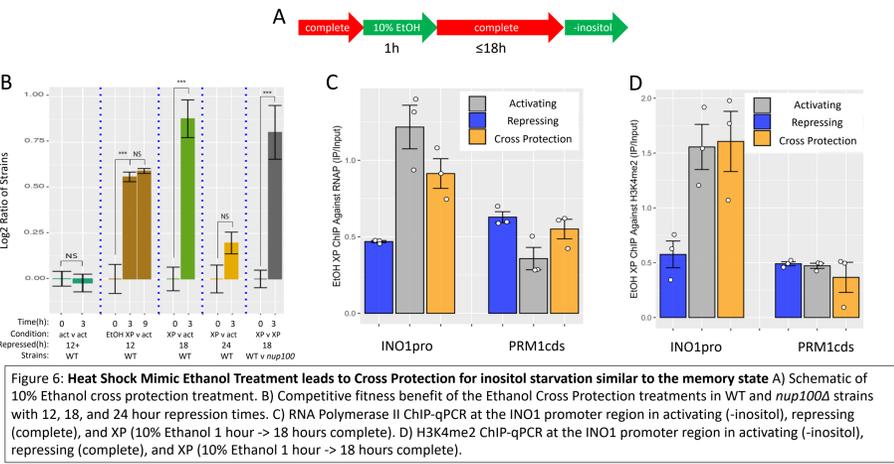


Figure 6: **Heat Shock Mimic Ethanol Treatment leads to Cross Protection for inositol starvation similar to the memory state** A) Schematic of 10% Ethanol cross protection treatment. B) Competitive fitness benefit of the Ethanol Cross Protection treatments in WT and *nup100Δ* strains with 12, 18, and 24 hour repression times. C) RNA Polymerase II ChIP-qPCR at the *INO1* promoter region in activating (-inositol), repressing (complete), and XP (10% Ethanol 1 hour -> 18 hours complete). D) H3K4me2 ChIP-qPCR at the *INO1* promoter region in activating (-inositol), repressing (complete), and XP (10% Ethanol 1 hour -> 18 hours complete).

The *INO1* gene experiences a phenomenon known as epigenetic transcriptional memory which is characterized by movement to the nuclear periphery, interaction with Nup100, a memory-specific chromatin state and PIC. This results in more rapid mRNA production and a competitive fitness benefit. The establishment and maintenance of these chromatin changes across generations are seen to be controlled by differing mechanisms and are self-perpetuating when induced at an ectopic site by incorporation of a Memory Recruitment Sequence. Components of this phenomenon are also seen when *INO1* is induced by a heat shock mimic showing variety in maintenance ability depending on how the gene is induced.